# Collaborative Research on the Northeast Water Polynya: NEWP92 Hydrographic Data Report USCGC *Polar Sea* Cruise, July 15 - August 15, 1992

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## **Sequence of Data Report Contents**

- Cruise description; Methods; Acknowledgements; References
- Station and Cast Listing
- Station Map
- Section Plots of Properties
- Vertical Profiles of CTD Properties
- CTD and Bottle Data Listings

#### **Introduction**

The Northeast Water Polynya (NEW) off the northeast coast of Greenland was the focus of two cruises aboard the USCGC *Polar Sea* during the summers of 1992 and 1993. The cruises were supported by the National Science Foundation Arctic Systems Science (ARCSS) program and were part of the Arctic Ocean Science Board's International Arctic Polynya Program. The Polar Sea cruises were designed as multidisciplinary studies to test hypotheses about the mechanisms of heat, water and carbon flow within and beyond the boundaries of the polynya. Preliminary results of the 1992 study have been described elsewhere (NEWATER, 1993). A collection of papers arising from the 1992 cruise have been published in a Special Section of the Journal of Geophysical Research (Overland et al., 1995).

This data report presents the hydrographic and basic chemical observations made from CTD/Rosette casts during the 1992 cruise. The station positions cruise are plotted in Figure 1. Also included in the report are selected section plots and vertical profiles. A total of 130 CTD casts were made during the cruise, measuring pressure, temperature, conductivity, dissolved oxygen, fluorescence and light transmission. Discrete samples were collected in 10-liter, rosettemounted, Niskin bottles and analyzed, from most casts, for: salinity, dissolved nutrients, dissolved oxygen, anthropogenic halocarbons (e.g., Freon gases), pigments, particulate organic carbon and nitrogen. Suspended particulate matter was analyzed at selected stations and these data were used to calibrate the CTD-transmissometer. Samples were collected from selected stations and depths for tritium and helium analyses, carbonate chemistry, as well as for measurements of bacterial abundance.

All the data listed in this report are currently stored in a remotely-accessible database at Brookhaven National Laboratory, access to which can be arranged on request (contact: D. Wallace). In addition, the entire NEWP92 and NEWP93 dataset is to be published in CD-ROM format by the National Snow and Ice Data Center, University of Colorado, Boulder, CO. The hydrographic data will also be submitted to NOAA's National Oceanographic Data Center.

#### **Description of Methods:**

#### *Header Data:*

Ice cover refers to bridge observations made by Coast Guard personnel at or close to the time of sampling. Specifically, ice cover at any particular station has been estimated from linear interpolation of the hourly bridge observations which were initially recorded in tenths. It should be noted that the ice cover in this region was often highly variable in time, as well as in space.

#### *CTD Data:*

CTD/Rosette Sampling. CTD data were collected using a Neil Brown Instrument Systems Mark III CTD underwater unit with a model 1150 deck data terminal. The CTD was equipped with a Beckman oxygen sensor and interfaced with a SeaTech fluorometer, and a 25cm path-length SeaTech transmissometer.

All underwater instrumentation was mounted on a General Oceanics rosette multi-bottle sampler, model 1015-12. The rosette was equipped with 11 x 10-liter Niskin bottles, and a single 5-liter bottom-tripped bottle. The bottom-trip switch was triggered when a weight suspende from a lanyard one meter beneath the rosette contacted the bottom.

CTD data were collected and displayed in real-time on a PC-486 computer using Oceansoft I software from EG&G Marine Instruments.

CTD Data Processing. The raw CTD data were transferred to a MicroVAX for processing. Large spikes were removed using a gradient check and 0.1 dbar averages were calculated. For each cast, data were extrapolated to the bottom depth. Any obvious large-amplitude spikes that remained in the raw data variables (pressure, temperature, conductivity, oxygen current, oxygen temperature, fluorometer and transmissometer) were edited and replaced by interpolation. Additionally, any missing values were filled in by linear interpolation.

To reduce the spikes caused in the calculation of salinity, a three-point moving average was first performed on conductivity to create a smooth signal. A lag of .4 was then applied to conductivity to better match the conductivity sensor with the temperature sensor. After final calibrations were applied to the CTD data, derived variables (depth, potential temp, salinity, oxygen in %-saturation and  $\mu$ mole kg<sup>-1</sup>, c<sub>p</sub>, SPM, sigma-t, sigma-p, sigma-1000, sigma-theta, Brunt-Vaisalla frequency, integrated density, steric height and freezing point) were calculate and then averaged using a pressure-centering procedure.

CTD Data Calibration. CTD data were calibrated solely with in situ methods and did not employ the use of pre- or post-cruise laboratory calibrations.

1. Pressure. A pressure offset of -0.5 dbar was applied to each CTD cast in its entirety. This correction accounted for the difference between the true and actual distance measured by the CTD pressure sensor at the surface of the water.

2. Temperature. CTD temperature was calibrated based on a linear regression with two in situ SIS reversing thermometers. A reiterative procedure produced the following relationship:

Corr T = 
$$
.996306 * CTD T + .002380
$$
,

which had a standard deviation of 0.007°C.

3. Conductivity. Final conductivity corrections were derived from a reiterative linear regression of upcast CTD conductivity and in situ bottle conductivity (inverted from salinity using corrected pressure and temperature). A pressure edit greater than 30 was used to eliminate bottles taken in the chlorophyll max layer. Time-series plots of water sample data minus CTD upcast conductivity differences were constructed to determine station subgroups. Three groups were identified, giving the following slope and bias terms, and their respective standard deviations:



4. Oxygen. Using the corrected pressure, salinity and temperature values, CTD values of oxygen were calculated from the upcast oxygen current and oxygen temperature sensors, following the algorithm of Owens and Millard (1985). Final oxygen coefficients were derived from reiterative linear regression of CTD upcast oxygen and in situ water sample oxygen, both in units of percent saturation. The same pressure edit criterion  $(P > 30)$  was used as stated above. Time-series plots yielded two groups, giving the following terms:

CTD 1-100 Corr Ox  $\% = .991460 * CTD$  Ox  $\% + .068328$  Std. dev. 0.0099 CTD 101-130 Corr Ox  $\% = .851967 * CTD$  Ox  $\% + .139818$  Std. dev. 0.0097

5. Transmissometer. Transmissometer data were calibrated by correlation with gravimetric analysis of suspended particulate loads. Between 1 and 10 liters of water were vacuum-filtered in-line from 10l water bottles through pre-weighed 0.4 µm Poretics PCTE membrane filters. The filters were rinsed in a laminar flow hood ten times with distilled, deionized and 0.4 µm filtered water, then air dried in the hood. At least one filter was put through this process during each cast without exposing it to seawater to establish an average blank value for methodological corrections. On deck transmissometer voltages were monitored throughout the cruise to assure clear optical pathways. Air calibrations of the transmissometer used were made in the lab at the beginning and end of the cruise to correct for any decay of the LED beam intensity. Readings with the light path blocked and unblocked were made through the CTD electronics prior to several casts to test for any voltage losses through the CTD. Percent transmission was recorded through the CTD and converted to a total beam attenuation coefficient (c) using the equation:

$$
V/5 = T = e^{-cz},\tag{1}
$$

where V is the instrument voltage output, 5 is the maximum voltage of the transmissometer output, T is percent transmission, c is the beam attenuation coefficients with units of  $m^{-1}$ , and z is the optical pathlength in meters. Thermal hysteresis was not corrected for because the thermal variability was small  $(\leq 5^{\circ}C)$  between the on-deck temperature and through the water column, and little or no hysteresis was apparent from comparison of up and down traces.

The beam attenuation coefficient in natural seawater is a summation of the beam attenuation coefficients for seawater  $(c_w$ , "yellow matter"  $(c_y)$ , and particles  $(c_p)$  (Pak et al. 1988):

$$
c = c_w + c_y + c_p. \tag{2}
$$

The contribution of  $C_w$  is constant and is set at the factory to equal 0.364 m<sup>-1</sup> in particle-free water for this instrument. At 660 nm,  $c_v$  is assumed constant and negligible (Bricaud et al., 1981; Gardner et al., 1993). To obtain  $c_p$ , c was calculated from the transmission values. The minimum value,  $c_{min}$ , for the cruise was subtracted from all c values to yield a pre-adjusted  $c_p$  (beam attenuation due to particles alone). Linear fits were performed between the pre-adjusted  $c_p$  values and the suspended particulate matter (SPM) concentrations obtained from the water filtered from the rosette water bottles. The data were split at a  $c_p = 0.25$ , essentially dividing the near surface data from the rest of the water-column. The x-intercept value from the  $c_p < 25$  relationship was subtracted from the pre-adjusted  $c_p$  to obtain a  $c_p$  that was zero for a particle mass

concentration of zero. The two linear fits were then equilibrated at  $c_p = 0.453$  to yield a single relationship between  $c_p$  and SPM concentration for the entire range of transmissometer data.

## *Bottle Data*

BNL ID: Each Niskin bottle closed during the cruise was assigned a unique identifier number that was incremented throughout the cruise; this same number was used to identify all subsamples collected from that particular Niskin. This 'BNL ID' number was subsequently used to merge data arising from various analytical procedures into a single database.

Dissolved Oxygen: Sub-samples for Winkler titrations were drawn into  $\sim$ 125 ml flasks immediately after the rosette was brought on deck and samples had been withdrawn for chlorofluorocarbon analyses. These samples were analyzed following the methodology described by Carpenter (1965). Subtle changes to methodology and calibration as described by Culberson (1991) were implemented, so that the oxygen data meet the precision and accuracy guidelines of the World Ocean Circulation Experiment (WOCE, 1991): namely accuracy <1% and precision  $-0.1\%$ .

Dissolved Nutrients: Sub-samples of ~60 ml were drawn from the Niskin bottles within 10-20 minutes of the cast being complete. Phosphate, silicate, nitrate, nitrite and ammonium were measured as soon as possible after sample collection (usually within a few hours) using a Technicon Autoanalyzer II, following standard colorimetric methods. The methods used have been described in Whitledge et al. (1981) with the exception of the phosphate determination which used the hydrazine reductant method described in Gordon et al. (1992). Standards were analyzed with each batch of samples in order to compensate for instrument response drift. Standards were prepared in both distilled, deionized water and low-nutrient surface seawater to determine the salt-effect on colorimeter response. The wash water was distilled, deionized water.

Total Dissolved Inorganic Carbon and Total Alkalinity: Samples were collected in 500 ml ground-glass stoppered bottles, to which 200  $\mu$ l of 50%-saturated HgCl<sub>2</sub>, was added. A headspace was introduced to allow for expansion, and the bottle was then tightly stoppered. The stopper was coated with Apiezon grease, and held firmly in place with elastic-bands. Samples were kept cool  $({}_{5}^{\circ}C)$  following collection and prior to analysis. Subsequently they were analyzed in the laboratory at Brookhaven National Laboratory for total dissolved inorganic carbon by coulometric titration (using a SOMMA system: Johnson and Wallace, 1992; Johnson et al., 1993). Small corrections were applied to account for the loss of  $CO<sub>2</sub>$  gas to the headspace, and for the dilution of the sample due to addition of  $HgCl<sub>2</sub>$ . Total alkalinity analyses were performed subsequently on the same samples at the Woods Hole Oceanographic Institution (Dr. Catherine Goyet's laboratory) using potentiometric titration in a closed, glass cell.

Pigments: Subsamples ( $\sim$ 280 ml) were filtered through Gelman GF/F glass fiber filters ( $\sim$ 0.7 µm nominal pore size). The filters were sonicated (on ice, in darkness) for 10 minutes in glass 15 ml centrifuge tubes together with 10 ml of 90% acetone. The samples were extracted in the dark for an additional 15 minutes and read using a Turner Designs Model 10 fluorometer before and after acidification (Smith and Nelson, 1990). The fluorometer was calibrated at the beginning and end

of the cruise using a commercially prepared chlorophyll standard.

Particulate Organic Carbon and Nitrogen: POC and PON were quantified by filtering samples onto precombusted (450°C for 2 hours) GF/F Whatman glass fiber filters, placing them in combusted glass vials, drying the filters at 60° C, and analyzing them on a Carlo-Erba model EA1108 elemental analyzer after high-temperature pyrolysis.

Tritium and Helium: For tritium analysis, 1-liter water samples were drawn from the Niskins into glass bottles, which had been previously filled with argon gas. These bottles were capped with a minimum of head-space and transferred to the laboratory. In the laboratory the samples were transferred to 1-liter Corning 1724 glass flasks in a vacuum line, degassed, sealed, and placed in a storage freezer at -20°C. After about 8 months, accumulated <sup>3</sup>He in the flask was measured in a MAP215-50 mass-spectrometer and the tritium concentration was calculated using calibrated air aliquots as standards, and appropriate correction factors. In addition, a set of NBS standards prepared to have 1 - 2 TU were measured as a check on accuracy. The reproducibility of the NBS standards and duplicate samples was  $0.007$  TU. Analytical precision was  $\sim$ 2.5% of the measurement or ~0.007 TU, whichever was greater (1 TU =  $1 \text{ }^3\text{He}/10^{18}\text{H}$ ). Measurements were decay-corrected to the time of sampling.

For helium analysis, 50 g water samples were sealed in refrigeration-grade copper tubes by clamping, and transferred to the laboratory. In the laboratory, water samples were processed in a vacuum line to seal the helium-neon fraction of the dissolved gas into Corning 1724 glass ampoules. These were then re-processed in the mass-spectrometer inlet line to separate helium from neon, and the helium fraction was admitted into the MAP 215-50 mass-spectrometer. <sup>3</sup>He and  ${}^{4}$ He beams were measured separately, and calibrated with standard air aliquots. The  ${}^{3}$ He ratio anomaly was expressed as:

$$
\delta^{3} \text{He} = \left[ \left[ \left( \frac{3}{4} \text{He} / 4 \text{He} \right)_{\text{sx}} / \left( \frac{3}{4} \text{He} / 4 \text{He} \right)_{\text{std}} \right] - 1 \right] \times 100,
$$

where sx and std refer to the sample and the atmospheric standard, respectively.  ${}^{4}$ He measurements were converted to concentrations by peak height comparison, and were expressed in units of nanomoles per kg (seawater). The precision in  $\delta^3$ He is  $\pm 0.2\%$ ; in <sup>4</sup>He concentration, it was  $\pm 1.0\%$ .

Bacterial Abundance. Water samples were collected aseptically directly from the Niskin bottles immediately after arrival of the CTD on deck. Volumes of 10 ml were fixed in 0.2-um filtered 2% formaldehyde and stored at 4°C in the dark until processed for counting. We used a dual staining procedure, reported by Deming et al. (1995), to enumerate bacteria. Samples were stained with acridine orange (AO), according to Hobbie et al. (1977), and gently filtered onto a 0.2-µm black Nucleopore filter, followed by DAPI stain (0.001%) according to Porter and Feig (1980). The slide preparations were viewed using a Zeiss epifluorescence microscope. Individual bacteria in 20 randomly selected fields (a minimum of 200 bacteria per slide) were counted routinely using standard optical filters for AO; switching to optical filters for DAPI allowed confirmation that AO-fluorescing particles were microorganisms.

Units: All concentration data are reported in units of per kg (seawater), with the exception of bacterial abundance which is in units of cells per ml.

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#### **NEWP92 STATION LIST**

 **STATION CAST CTD-NO DATE TIME LATITUDE LONGITUDE SONIC DEPTH GMT GMT DEG. DEG. METERS** 1 | 1 | 1 | 7/18/92 | 08:42 | 76.526 | -6.907 | 1343 1 2 2 7/18/92 11:15 76.512 -6.966 1304 2 3 3 7/19/92 00:28 77.155 -10.481 488 2 4 4 7/19/92 01:49 77.160 -10.447 476 2 5 5 7/19/92 03:01 77.160 -10.406 472 2 6 6 7/19/92 11:21 77.150 -10.449 487 3 7 7 7 7/20/92 12:43 78.606 -13.487 142 3 8 8 7/20/92 14:17 78.609 -13.508 139 3 9 9 7/20/92 15:28 78.616 -13.528 137 3 10 10 7/20/92 19:49 78.645 -13.494 168 4 | 11 | 11 | 7/21/92 | 13:55 | 80.398 | -14.243 | 337 4 | 12 | 12 | 7/21/92 | 15:11 | 80.410 | -14.208 | 333 4 | 13 | 13 | 7/21/92 | 16:29 | 80.421 | -14.156 | 330 5 14 14 7/22/92 13:12 80.652 -11.269 238 6 | 15 | 15 | 7/22/92 | 15:51 | 80.649 | -12.169 | 260 7 16 16 7/22/92 19:57 80.648 -13.082 222 8 17 17 7/24/92 02:29 80.532 -14.661 208 9 | 18 | 18 | 7/24/92 | 05:22 | 80.483 | -14.488 | 324 10 | 19 | 19 | 7/24/92 | 07:03 | 80.440 | -14.422 | 301 11 | 20 | 20 | 7/24/92 | 08:16 | 80.369 | -14.088 | 314 12 21 21 7/24/92 10:33 80.307 -13.797 286 13 22 22 7/24/92 12:22 80.238 -13.617 175 14 23 23 7/24/92 13:39 80.188 -13.454 185 15 24 24 7/24/92 16:01 80.119 -13.159 75 16 25 25 7/25/92 08:33 80.275 -11.029 199 17 | 26 | 26 | 7/25/92 | 11:13 | 80.189 | -10.923 | 121 18 27 27 27 7/25/92 13:44 80.338 -11.026 253 19 28 28 7/25/92 15:31 80.409 -11.031 315 20 29 29 7/25/92 17:43 80.477 -10.897 290 21 30 30 7/25/92 19:44 80.539 -10.981 257 22 31 31 7/25/92 21:16 80.609 -10.975 267 23 32 32 7/25/92 22:17 80.671 -10.956 181 24 | 33 | 33 | 7/25/92 | 23:41 | 80.722 | -10.938 | 111 25 34 34 7/26/92 08:37 80.443 -13.332 288 25 35 35 7/26/92 14:12 80.450 -13.349 306 25 36 36 7/26/92 20:58 80.455 -13.401 305 25 37 37 7/27/92 03:16 80.454 -13.328 301 25 38 38 7/27/92 03:51 80.458 -13.302 301 25 39 39 7/27/92 04:43 80.464 -13.272 302



